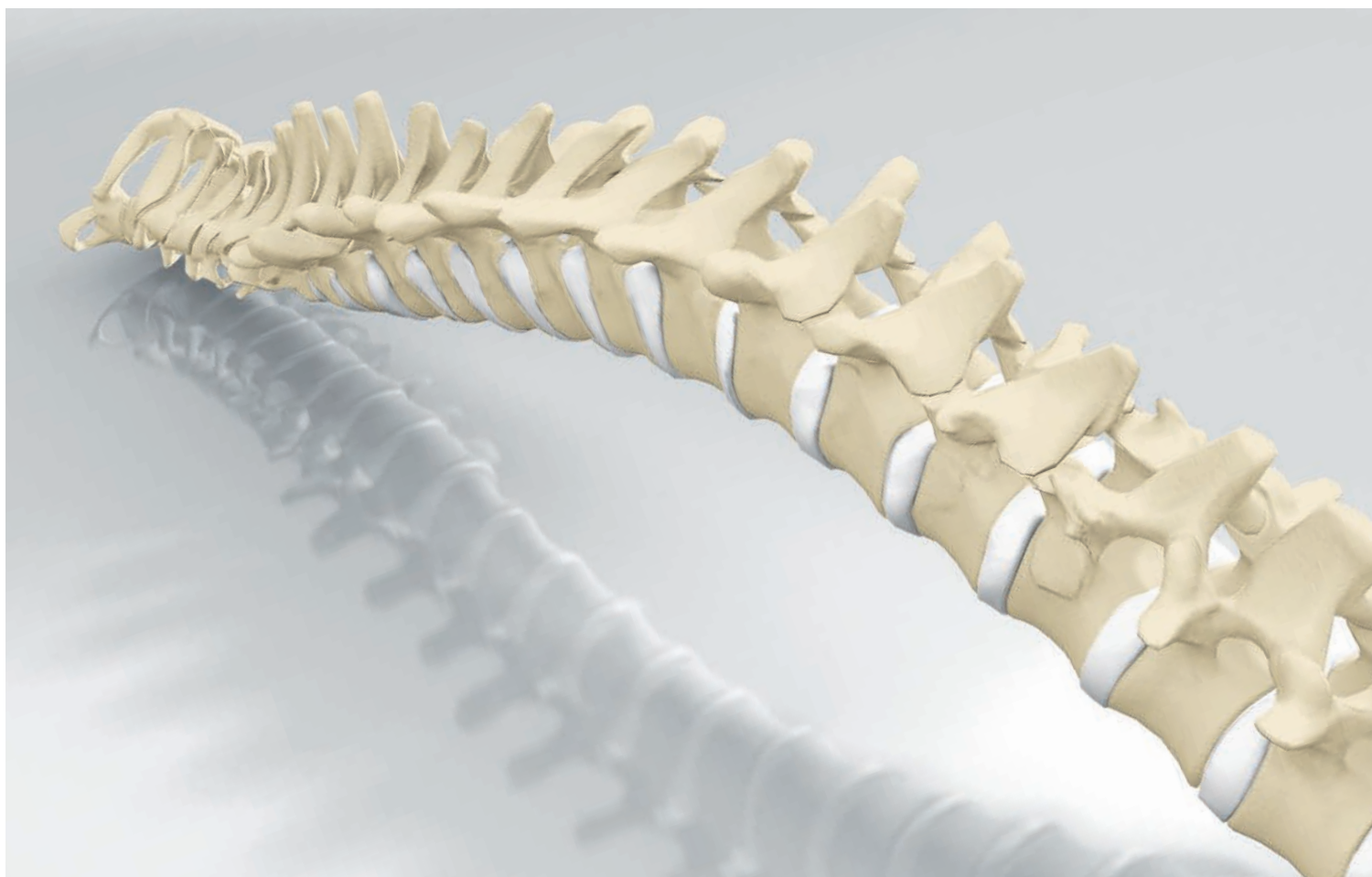




SPINE SURGERY

AESCULAP[®] CeSPACE[®] 3D
CERVICAL INTERBODY FUSION SYSTEM
SURGICAL MANUAL

AESCULAP® CERVICAL SPINE



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AESCULAP® CeSPACE® 3D

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PROTECTING AND PRESERVING SPINAL STABILITY

Modern lifestyle has resulted in increasing physical inactivity among people all over the world. Of the many medical problems associated with this, spinal disorders are among the most critical. This is even more significant as the spinal column is one of the most important structures in the human body. It supports and stabilizes the upper body and is the center of our musculoskeletal system, which gives the body movement. Our work in the field of spine surgery is dedicated to protecting the spinal column and preserving its stability. We support spine surgeons with durable, reliable products and partner services for reliable procedures and good clinical outcomes (1-10).

Our philosophy of sharing expertise with healthcare professionals and patients allows us to develop innovative implant and instrument systems that help to preserve stability and stabilize the cervical and thoracolumbar spine.

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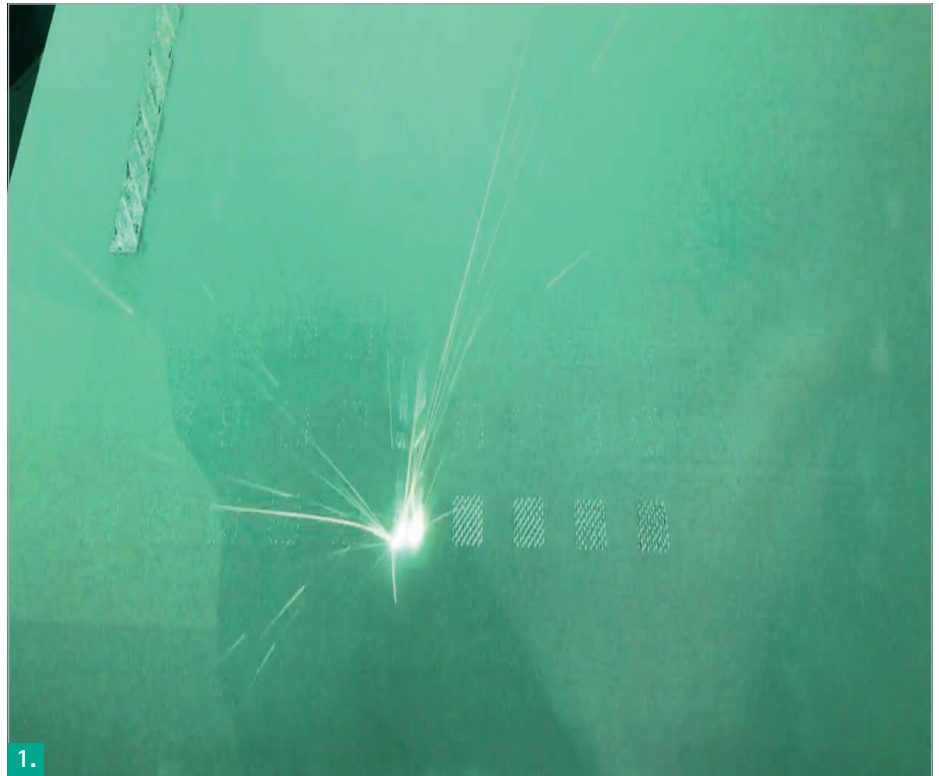
A. IMPLANT MATERIAL

THE TECHNOLOGY OF LASER SINTERING – A WELL-ESTABLISHED ADDITIVE LAYER BY LAYER PROCESS

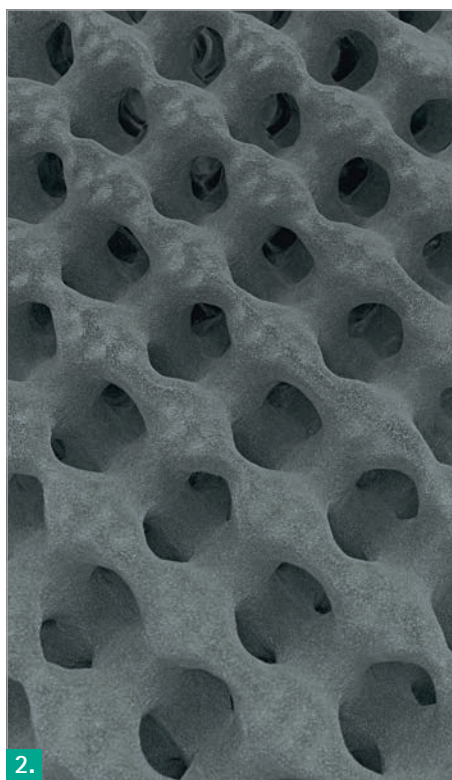
■ Additive manufacturing – 3D printing – means a layer by layer process to design a device using laser beam and metal powder. This innovative laser beam melting technology is of growing importance in the manufacture of implants, as it allows to create various fine and porous surface structures with the aim to support bone-ingrowth. Homogenous or heterogeneous lattice structures or combinations of various kinds of structures and surfaces are generally conceivable.

- Direct assembly of the component based on 3D-CAD data
- Design freedom

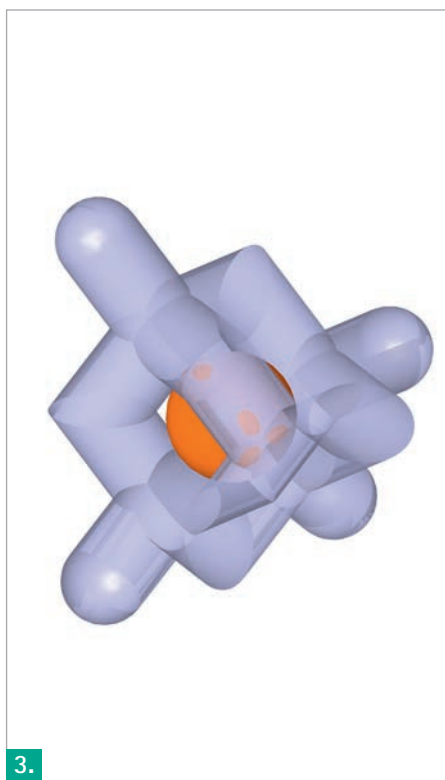
We combined our long-time experience in designing and manufacturing spinal implants with latest technology and produce in-house our AESCULAP® 3D Cages (Fig. 1).



Laser beam melting technology



2. Lattice structure Structan®



3. Unit cell with fitted ball of 900 µm

AESCULAP® 3D Cages are engineered from Structan® – a lattice structure with largely isotropic behavior. Ti6Al4V ELI was chosen as a proven and biocompatible material for implants (11).

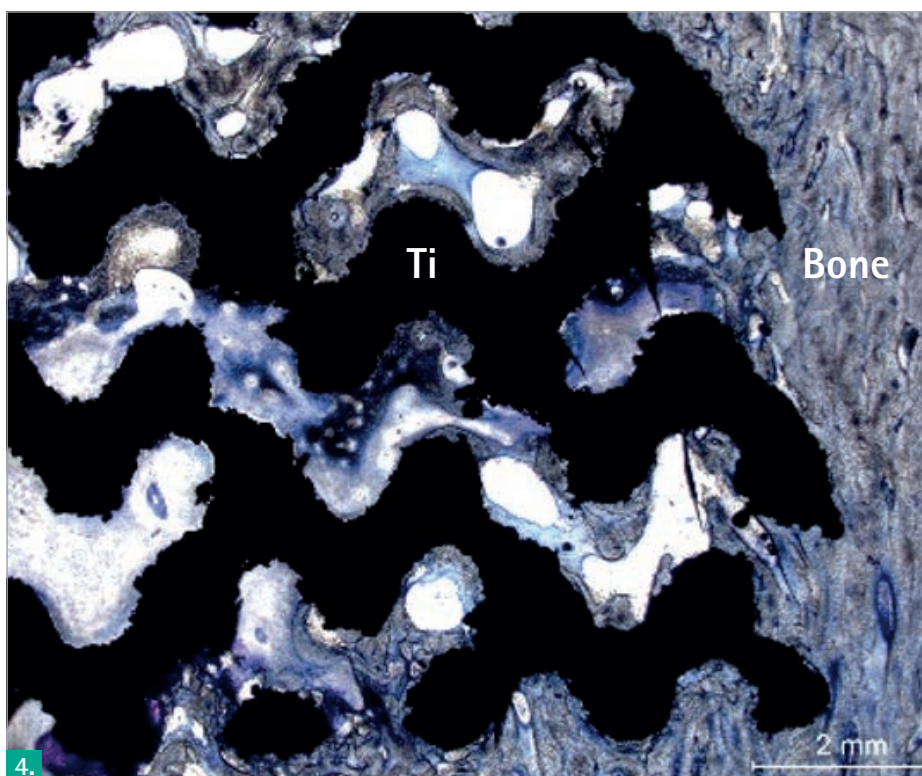
MORE CONNECTION

■ The lattice structure of the AESCULAP® 3D Cages shows an interconnected pore structure (Fig. 2/3). This interconnectivity facilitates migration of bone cells into the structure, thereby providing secondary stability (12, 13).

■ According to the average pore size and porosity of cancellous bone (approximately 1 mm/50-90% (14)) the 3D lattice structure Structan® features an all-over regular pore size of 900 µm as well as a mean interconnected porosity of 50-55%. Pore size and porosity are in a favorable range to stimulate bone in-growth (15, 16).

■ The results of a sheep study with partly loaded implants confirm bone growth on and into the 3D lattice structure without connective tissue layer six months post-operatively. This formation of bone tissue within the 3D lattice structure leads to a high secondary stability (13). The 3D lattice structure serves as a guide rail for bony integration and thus contributes significantly to the secure anchoring of the 3D Cage (Fig. 4).

■ A rough laser sintered surface provides a good interaction between bone cells and implant surface compared to a milled smooth surface and therefore intends to optimize osseointegration (17).



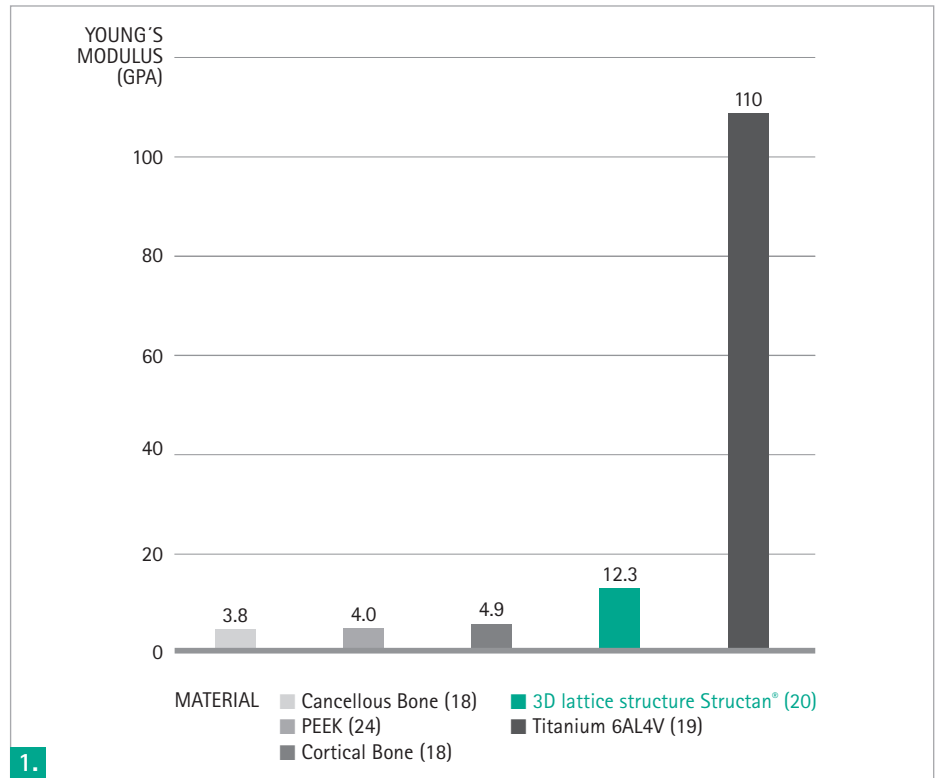
4. Histological section of the 3D Cage lattice structure filled with newly formed bone

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A. IMPLANT MATERIAL

MORE ELASTICITY

Ti6Al4V ELI as solid material has a Young's modulus of approximately 110 GPa as it is shown in the figure, whereas cortical bone has a Young's modulus of approximately 5 GPa (18, 19). The Young's modulus of Structan® is developed to be close to that of cortical bone (20). This may prevent subsidence into the vertebral body (21). In addition, this may result in improved bone growth (22) (Fig. 1).

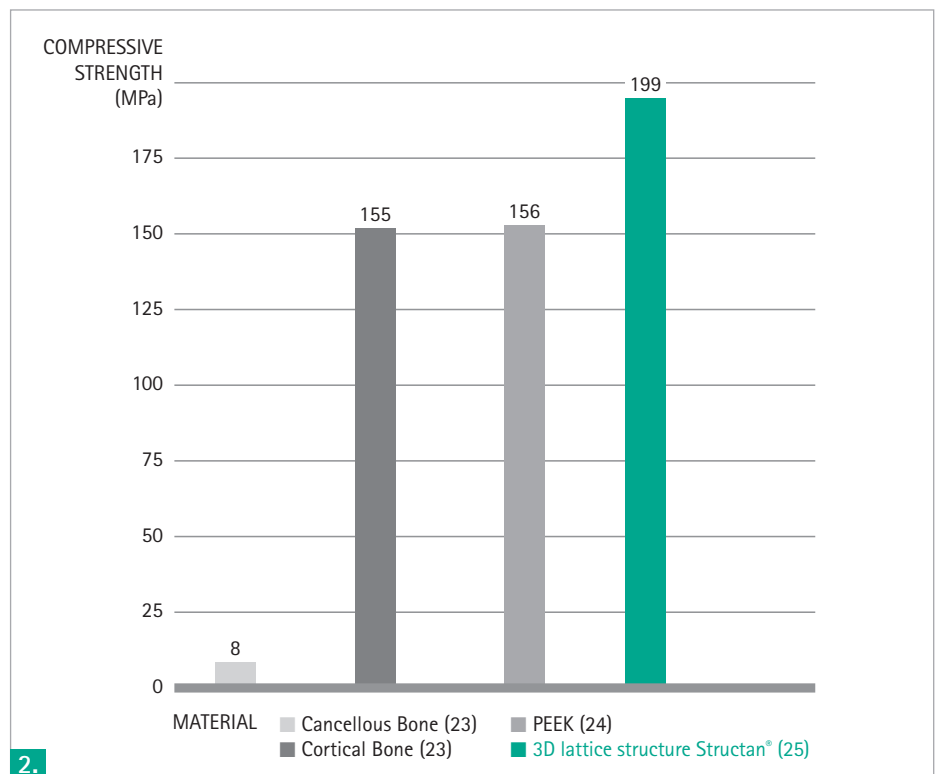


1.

Young's modulus of various materials

MORE STRENGTH

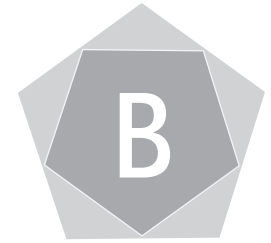
The 3D lattice structure Structan® combines a bone-like Young's modulus with a high compressive strength, which contributes to high safety against failure due to breakage. The compressive strength of the 3D lattice structure Structan® is higher than the mean strength of bone and PEEK (23-25) (Fig. 2).



2.

Compressive strength of 3D lattice structure Structan®

B. INTENDED USE & IMPLANT DESIGN



■ Stabilization of the cervical spine C2-T1 through anterior approach, monosegmental and multisegmental.

■ A cervical plate may be required for additional stabilization.

■ Solid frame without sharp edges for biomechanical stability and smooth insertion into the disc space minimizing the risk to injure surrounding soft tissue.

■ Open porous structure designed to provide primary and secondary stability.

■ The implant's anatomical endplate design provides a good contact area between implant and vertebral endplates whilst allowing addition of bone material to enable bone growth through the center of the implant.

■ Cranial und caudal anchoring elements in form of spikes for a firm implant fit and high primary stability.

■ Trapezoidal shape intended to fit anatomical conditions.

■ Screw thread interface allows a firm connection to inserter.

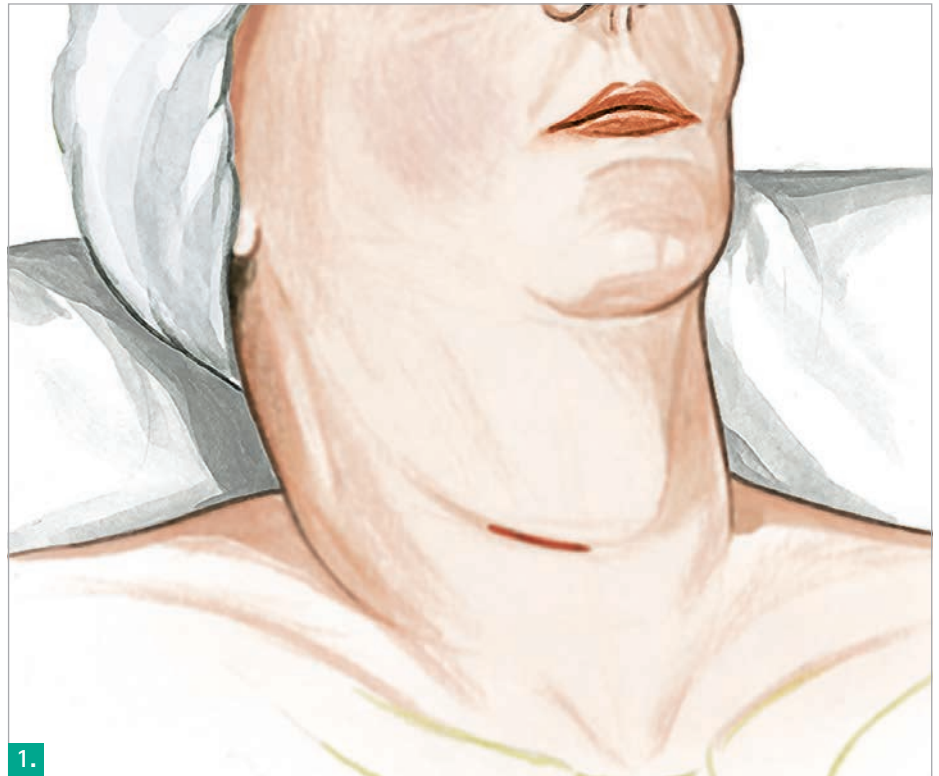
■ Good visibility in X-ray to localize implant positioning (26, 27).

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C. SURGICAL TECHNIQUE

C.01. PATIENT POSITIONING

■ The patient is placed in the supine position with the head slightly reclined and stabilized in a head holder (Fig. 1). Once the lordotic cervical spine has been supported, the thorax may be placed on a pillow to emphasize the reclinatio



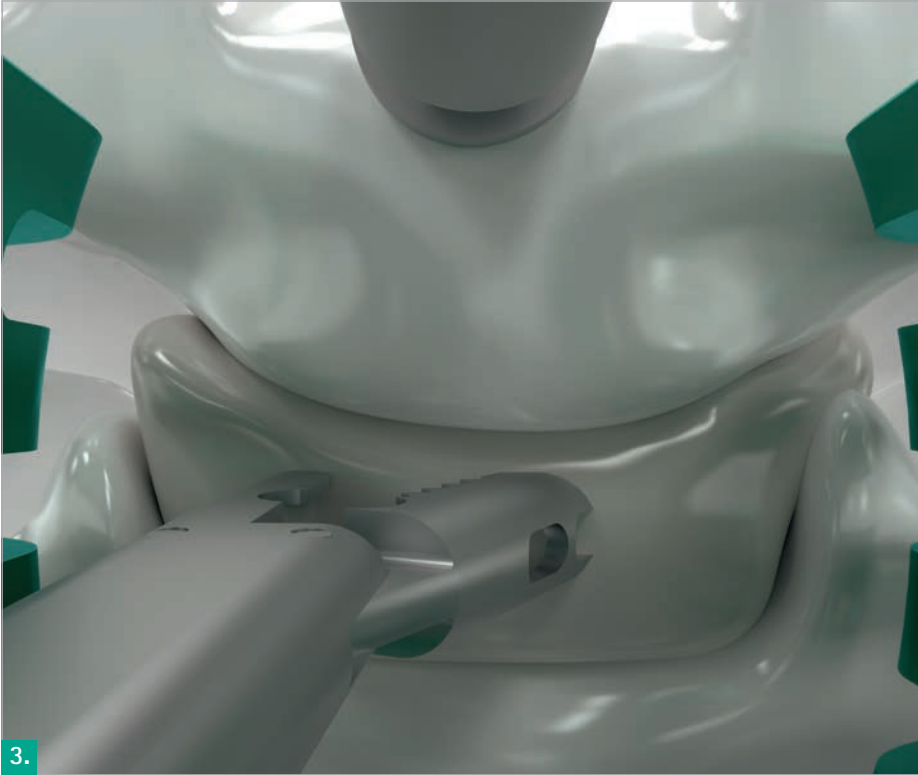
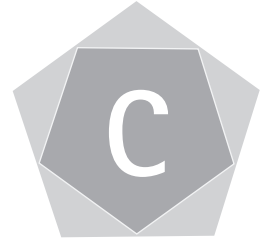
C.02. EXPOSURE OF THE INTERVERTEBRAL SPACE

■ After the skin incision and preparation, the CCR retractor is placed. The blades are available in PEEK and titanium (Fig. 2). A counter retractor can be used. The subcutaneous tissue is separated from the platysma cranially, caudally and medially, and the platysma is also separated following the direction of its fibres. The margins of the platysma can be held apart with the retractor or with two surgical forceps.

■ Now the medial edge of the sternocleidomastoid muscle is located and prepared with the index finger in the connective tissue space over the ventral surface of the cervical spine and under lateralization of the vascular nerve bundle and medialization of the trachea, esophagus and thyroid gland.

■ After the Langenbeck hooks have been inserted, the ventral surface of the cervical spine, still covered by a thin prevertebral layer of connective tissue, is revealed. This layer can now be exposed by either a blunt scissor or alternatively through bipolar coagulation, in order to expand the tissue cranially and caudally using a swab. A wire is set under X-ray monitoring to mark the intervertebral disc space.





C.03. DISTRACTION / DISCECTOMY / PREPARATION OF THE ENDPLATES

- The distraction screws are placed in position and the CASPAR® distractor is applied following the CASPAR® technique (Fig. 2). Complete discectomy is performed using various rongeurs, rectangular curettes and bone curettes (Fig. 3). While using a high speed drill to remove the posterior rim and/or dorsal osteophytes, care must be taken to avoid damaging the vertebral body endplates.

PLEASE NOTE

Make certain that the endplates of the neighboring vertebral bodies are not weakened, in order to minimize the risk of migration.

Make certain that the implant bed is properly prepared to avoid damage to the implant when it is driven in.

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C. SURGICAL TECHNIQUE

C.04. ASSEMBLING OF THE TRIAL IMPLANTS

- The depth stop is snapped onto the shaft of the trial implant (Fig. 1).
- Markings on the depth stop indicate the correct assembling.

INFORMATION

Use CeSPACE® 3D trial implant with depth stop.



C.05. IMPLANT SELECTION

- Use trial implants to establish the correct implant size.
- When inserting the trial implant observe the markings to correctly align the trial implant. Markings on the handle as well as on the trial itself indicate the cranial side of the trial (Fig. 2/3).

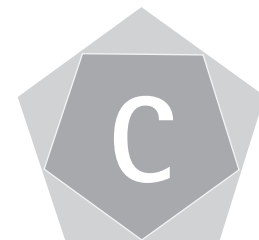
DETERMINATION OF IMPLANT SIZE

- The CeSPACE® 3D trials have the anatomical shape of the CeSPACE® 3D implants.

PLEASE NOTE

The trials are essential to ensure the correct implant size to be used.





C.06. IMPLANT REMOVAL FROM PACKAGING

- Open folder blister to remove the CeSPACE® 3D implant.
- The packaging concept allows implant removal with the connected inserter.

C.07. ASSEMBLING OF THE INSERTION INSTRUMENT

- The depth stop is snapped onto the shaft of the insertion instrument. It prevents the implant from being inserted too deeply into the intervertebral disc compartment (Fig. 4).

PLEASE NOTE

Use CeSPACE® 3D insertion instrument with depth stop.



C.08. FILLING OF CAGE

- Use the packing block and the punch for optional filling of the implant with bone or bone substitute (Fig. 5).
- The CeSPACE® 3D implant is connected with the inserter.

PLEASE NOTE

Do not use force during filling to avoid implant damaging.

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C. SURGICAL TECHNIQUE

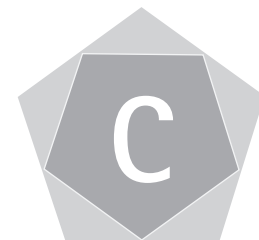
C.09. CeSPACE[®] 3D INSERTION

- The CeSPACE[®] 3D implant is held securely and firmly onto the CeSPACE[®] 3D inserter by means of a screw joint (Fig. 1).
- Once CeSPACE[®] 3D is attached to the inserter, it can be introduced into the intervertebral space using image converter monitoring (Fig. 2).
- When inserting the implant into the intervertebral space, observe the markings to correctly align the implant.

PLEASE NOTE

Implant marking points in the cranial direction.





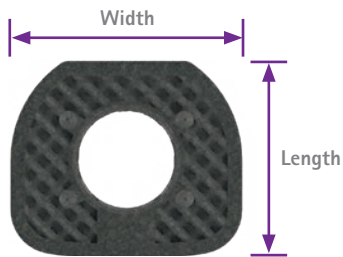
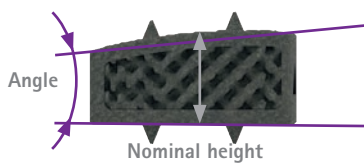
C.10. FINAL POSITIONING

- The implant should be inserted centrally in AP and with a distance of approximately 1 - 2 mm to both the anterior and posterior rim (Fig. 3).
- For additional stabilization, a cervical plate may be necessary.

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D. IMPLANT OVERVIEW

LORDOSIS 5°





Article No.	Size (Height x Width x Length)	Quantity
SN634T	4 x 14 x 11.5 mm	2
SN635T	5 x 14 x 11.5 mm	2
SN636T	6 x 14 x 11.5 mm	2
SN637T	7 x 14 x 11.5 mm	2
SN638T	8 x 14 x 11.5 mm	2
SN644T	4 x 16 x 13.5 mm	2
SN645T	5 x 16 x 13.5 mm	2
SN646T	6 x 16 x 13.5 mm	2
SN647T	7 x 16 x 13.5 mm	2
SN648T	8 x 16 x 13.5 mm	2
SN654T	4 x 18 x 15 mm	2
SN655T	5 x 18 x 15 mm	2
SN656T	6 x 18 x 15 mm	2
SN657T	7 x 18 x 15 mm	2
SN658T	8 x 18 x 15 mm	2

E. INSTRUMENT OVERVIEW








SN600 – CeSPACE® 3D INSTRUMENTATION

	Article No.	Description	Quantity
	SN601R	CeSPACE® 3D tray f/instrumentation	1
	TF353	Packing stencil f/SN601R (SN600)	1
	JA455R	Lid for OrthoTray DIN w/o handle	1
	SN664R	CeSPACE® 3D trial implant 5°, 4 x 14 x 11.5 mm	1
	SN665R	CeSPACE® 3D trial implant 5°, 5 x 14 x 11.5 mm	1
	SN666R	CeSPACE® 3D trial implant 5°, 6 x 14 x 11.5 mm	1
	SN667R	CeSPACE® 3D trial implant 5°, 7 x 14 x 11.5 mm	1
	SN668R	CeSPACE® 3D trial implant 5°, 8 x 14 x 11.5 mm	1
	SN674R	CeSPACE® 3D trial implant 5°, 4 x 16 x 13.5 mm	1
	SN675R	CeSPACE® 3D trial implant 5°, 5 x 16 x 13.5 mm	1
	SN676R	CeSPACE® 3D trial implant 5°, 6 x 16 x 13.5 mm	1
	SN677R	CeSPACE® 3D trial implant 5°, 7 x 16 x 13.5 mm	1
	SN678R	CeSPACE® 3D trial implant 5°, 8 x 16 x 13.5 mm	1

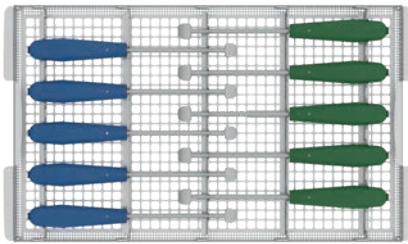
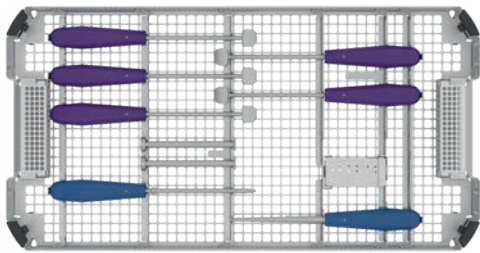
AESCULAP® CeSPACE® 3D

E. INSTRUMENT OVERVIEW

SN600 – CeSPACE® 3D INSTRUMENTATION

	Article No.	Description	Quantity
	SN684R	CeSPACE® 3D trial implant 5°, 4 x 18 x 15 mm	1
	SN685R	CeSPACE® 3D trial implant 5°, 5 x 18 x 15 mm	1
	SN686R	CeSPACE® 3D trial implant 5°, 6 x 18 x 15 mm	1
	SN687R	CeSPACE® 3D trial implant 5°, 7 x 18 x 15 mm	1
	SN688R	CeSPACE® 3D trial implant 5°, 8 x 18 x 15 mm	1
	SN663R	CeSPACE® 3D depth stop	2
	SN604R	CeSPACE® 3D packing block	1
	FF913R	CASPAR® graft positioning tamp – 3 mm diameter	1
	SN605R	CeSPACE® 3D insertion instrument	1

Recommended container: JK441
 Recommended container lid: JK489
 Recommended identification label: JG790B



SN600

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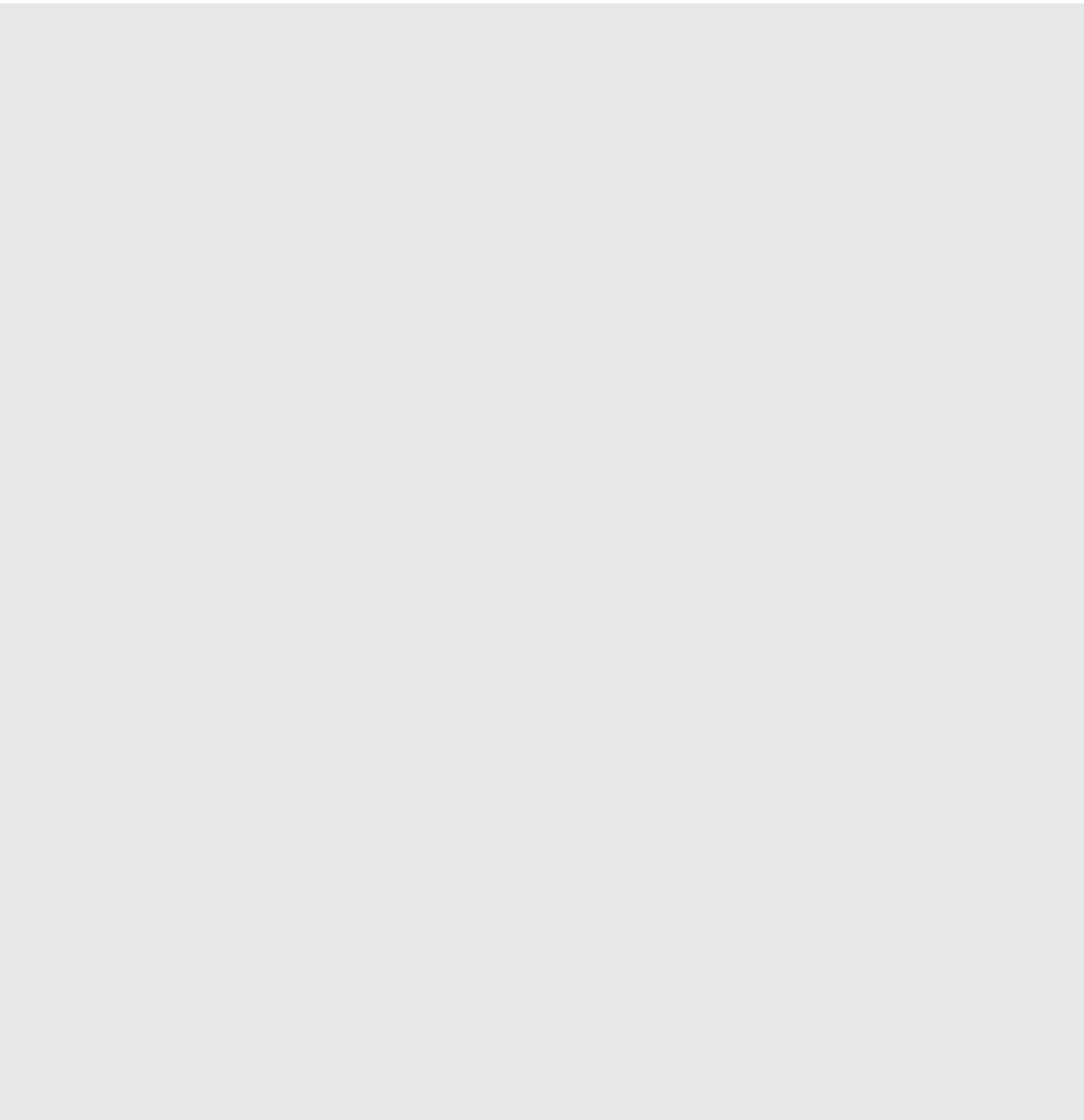
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The usability of the AESFULAP[®] 3D Cage System CeSPACE[®] 3D was tested in April 2019, in a cadaver workshop with six independent test persons as intended users (surgeons specialized in spinal surgery or comparable fields). Parameters such as implant visibility under x-ray control, mechanical stability of the implant/instrument interface and implant surface evaluation in terms of tissue injury risk were tested among others. Acceptance criteria were fulfilled for all the above-mentioned parameters. All test users confirmed the absence of critical features that must be improved prior to clinical use. During the test, the x-ray visibility of the cages was particularly positively assessed.

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CT and X-ray visualization of different AESFULAP[®] interbody fusion cages (full titanium, porous Ti6Al4V and PLASMAPORE^{XP}[®] cages) was tested in a cadaver setup. A radiologist evaluated the implant visibility and the presence of artefacts that may limit the visualization of adjacent structures. Visualization and assessment of implant position was achieved in X-ray and CT for all tested cages. Minor artefacts were visible in CT reconstructions in the surrounding of porous Ti6Al4V and full titanium implants. Porous Ti6Al4V implants showed slightly fewer artefacts in CT in comparison to full titanium implants. The minor artefacts observed did not limit the assessment of the surrounding anatomical structures.



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